CBCS: 2024-25 [NEP 2020]

F. Y. B.Sc.- Microbiology





SAVITRIBAI PHULE PUNE UNIVERSITY

(Formerly University of Pune)

Four Year B.Sc. Degree Program in Microbiology

(Faculty of Science and Technology)

Choice-Based Credit System Syllabus (2024 Pattern)

(As Per NEP 2020)

First Year B. Sc

To be implemented from Academic Year 2024-2025

Prepared by: B.O.S. MICROBIOLOGY, SPPU

Recommended by: Faculty, Science and Technology

Approved by: Academic Council, SPPU

(For Colleges Affiliated to Savitribai Phule Pune University, Pune)

Title of the Program: B.Sc. (Microbiology)

Preamble:

Microbiology is a wide-ranging discipline of biology. It covers five major groups of microorganisms i.e., bacteria, protozoa, algae, fungi, viruses. In this subject, the interactions of these microorganisms with their surroundings are studied systematically. This also focuses on how the potential of these organisms can be tapped in improving human life and their impact on society and civilization. Being a branch of biology, microbes are used as a study model in different conventional and modern areas of biology. Microbiology has a great legacy of active research in pure and applied science from its establishment as a separate subject. Though the microorganisms were discovered by human over five hundred years ago, there are huge opportunities to explore their wide diversity. In the changing scenario of the world and environment, systematic knowledge of microbiology has become quintessential and crucial. There is always a demand for skilled and knowledgeable persons in education, research and industry. Students who graduate by taking this subject as a major are employable in industry and research.

Introduction: In the post-globalization world, higher education has to play a significant role in the creation of skilled human resources for the well-being of humanity and the environment. The barriers among the academic fields seem to have dissolved. However, disparities in the field of curriculum aspect, evaluation and mobility exist. With the changing situations at local and global levels, the syllabus restructuring should keep pace with developments in the education sector. National Education Policy (NEP) is being adopted and implemented to address the issues related to the traditional system and it also aims to maintain the best of the earlier curriculum. The student is at the centre of NEP-2020. The present curriculum focuses on students' needs, skill development, interdisciplinary approach to learning and enhancing employability. Microbiology curricula are offered at two levels viz. undergraduate and postgraduate. The undergraduate curricula are prepared to impart basic knowledge of the respective subject from all possible angles. In addition, students are to be trained to apply this knowledge in day-to-day applications and to get a glimpse of research.

Objectives to be achieved:

- To enrich students' knowledge and train them in the pure microbial sciences
- To introduce the concepts of application and research in Microbiology

- To inculcate a sense of scientific responsibility and social and environmental awareness
- To help students build up a progressive and successful career

Eligibility for Admission:

First Year B.Sc.:

 a. Higher Secondary School Certificate (10+2) or its equivalent Examination with English and Biology; and two of the science subjects such as Physics, Chemistry, Mathematics, Geography, Geology, etc.

OR

b. Three Years Diploma in Pharmacy Course of Board of Technical Education conducted by Government of Maharashtra or its equivalent.

OR

c. Higher Secondary School Certificate (10+2) Examination with English and vocational subject of

+ 2 level (MCVC) - Medical Lab. Technician (Subject Code = P1/P2/P3)

Admissions will be given as per the selection procedure / policies adopted by the respective college keeping in accordance with conditions laid down by the University of Pune.

Reservation and relaxation will be as per the Government rules.

Medium of Instruction: English

Board of Studies (BoS) in Microbiology

From 2023-24 to 27-28

| Sr. No. | Name | Designation |
|---------|--------------------------------|-------------|
| 1. | Dr. Pawar Sunil Trimbak | Chairman |
| 2. | Dr. Pardesi Karishma Rajendra | Member |
| 3. | Dr. Pabale Anupama Ashok | Member |
| 4. | Dr. Wagh Pratima Pandit | Member |
| 5. | Dr. Abhyankar Pragati Sunil | Member |
| 6 | Dr. Pathak Leena Pradeep | Member |
| 7 | Dr. Kulkarni Snehal V. | Member |
| 8. | Dr. Kale Avinash Sudhakar | Member |
| 9. | Dr. Marathe Rajendra Jagannath | Member |
| 10. | Dr. Puranik Pravin R. | Member |
| 11 | Dr. Rajwade Jyotika Milind | Member |
| 12. | Dr.Mali Gajanan Vishnu | Member |
| 13 | Dr. Shete Ashiwini Monish | Member |
| 14 | Dr. Patil Hemant jagatrao | Member |

| Semester/Level | Course Type | Course Code | Theory/Practical | Title/Course Name | Credits |
|----------------|----------------------------|-------------|------------------|--|------------|
| SEM-I/4.5 | Subject 1 | MB-101-T | Theory | Introduction to Microbiology | 2 |
| | Subject 1 | MB-101-P | Practical | Laboratory Techniques in Microbiology-I | 2 |
| | Subject 2 | - | Theory | - | 2 |
| | Subject 2 | - | Practical | - | 2 |
| | Subject 3 | - | Theory | - | 2 |
| | Subject 3 | - | Practical | - | 2 |
| | Open Elective | OE-101-MB | Theory | Microorganisms and Human Life | 2 |
| | Skill enhancement Course | SEC-101-MB | Practical | Basic Skills in Microbiology Laboratory –I | 2 |
| | Indian Knowledge System | IKS-100-T | Theory | IKS Generic | 2 |
| | Ability Enhancement Course | AEC-101-MB | Theory | English/Hindi/Marathi | 2 |
| | Value Education Course | VEC-101-MB | Theory | Environmental science | 2 |
| | | | | | Total (22) |
| SEM-II/4.5 | Subject 1 | MB-151- T | Theory | Basic Microbiology | 2 |
| | Subject 1 | MB-151-P | Practical | Laboratory Techniques in Microbiology-II | 2 |
| | Subject 2 | - | Theory | - | 2 |
| | Subject 2 | - | Practical | - | 2 |
| | Subject 3 | - | Theory | - | 2 |
| | Subject 3 | - | Practical | - | 2 |
| | Open Elective | OE -151-MB | Practical | Dealing With Microorganisms | 2 |
| | Skill enhancement Course | SEC- 151-MB | Practical | Basic Skills in Microbiology Laboratory-II | 2 |
| | Ability Enhancement Course | AEC-151-MB | Theory | English/Hindi/Marathi | 2 |
| | Value Education Course | VEC-151-MB | Theory | Environmental science | 2 |
| | Certificate course | CC-151-MB | - | PE/NSS/NCC | 2 |
| | | | | | Total (22 |

Credit framework for First Year (UG)

Exit options, Award of credits, Evaluation pattern and ATKT rules are as per the SPPU

External Students

There shall be no external students.

University Terms

Dates for the commencement and the conclusion of the first and second terms will be declared by the University authorities. Terms can be kept by only duly admitted students. The term shall be granted only on a minimum of 80 percent attendance at theory and practical courses and satisfactory performance during the term.

Current curriculum orientation

To accommodate more advanced topics in the syllabi, it is necessary to understand the basic science knowledge level of the students who have chosen the microbiology discipline. Curricula of courses of state and central boards of higher secondary level were reviewed to avoid reiterations of previous syllabi.

Qualification of Teachers:

With a minimum undergraduate and postgraduate degree in microbiology (B.Sc. and M.Sc. Microbiology) and qualified as per UGC regulations.

Semester I

Paper I

MB-101-T: Introduction to Microbiology Total: 2 Credits Work-load: - 15 hrs/credit

(Total Workload: 2 credits \times 15 hrs =30 hrs in semester)

| | Course Objectives |
|---|--|
| 1 | To enrich the knowledge of undergraduate science faculty students about the |
| | different areas of microbiology. |
| 2 | To allow students to understand the mysterious world of microorganisms. |
| 3 | To explain the various categories of microorganisms and their general |
| | characteristics. |
| 4 | To make students understand the role of beneficial microorganisms present in different habitats. |
| 5 | To allow students to understand the general classification scheme of living things. |
| 6 | To understand the historical developments in the field of microbiology. |
| 7 | To enrich students' knowledge about recent inventions and discoveries in microbiology. |

| | Course Outcomes (COs) On completion of the course, the students will be able to: | |
|-----|--|--|
| CO1 | Describe the origin of microbial life on earth by outlining the experiment relating to | |
| | the formation of organic matter in a laboratory set-up. | |
| CO2 | Explain the evolution of the microbiology field and the scientific discoveries | |
| | relating to each field. | |
| CO3 | Outline the relatedness of the different upcoming areas of biological sciences to the | |
| | field of microbiology. | |
| CO4 | Classify different species according to the different classification systems. | |
| CO5 | Review the differences between the basic types of cells found in all biological | |
| | systems. | |
| CO6 | Write about the morphological and differential characteristics of different | |
| | groups of microorganisms. | |

| Credit | | Unit/ Topic details | No. of |
|--------|--------|--|--------|
| no. | | | hours |
| Ι | Devel | lopment of microbiology | 15 |
| | 1. | Discovery of Microscope and Microorganisms | 1 |
| | • | Anton von Leeuwenhoek | |
| | • | Robert Hooke | |
| | 2. | Abiogenesis v/s biogenesis | 2 |
| | • | Aristotle's notion about spontaneous generation, | |
| | • | Francesco Redi's experiment | |
| 1 | 3. | Golden Era of Microbiology | 4 |
| | • | Louis Pasteur (Rabies, Pasteurization) | |
| | • | Robert Koch (Koch's Postulates, Germ theory of disease) | |
| | • | Discovery of viruses (TMV) | |
| | • | Paul Ehrlich (Chemotherapy) | |
| | • | Edward Jenner (Vaccination) | |
| | • | Alexander Fleming (Penicillin) | |
| | 4. | Modern Era of Microbiology- | 3 |
| | Carl W | Voese classification based on 16S rRNA. | |
| | Signif | icance and application of Human Microbiome. | |
| | Nano- | biotechnology and Space Microbiology. | |
| | 5. | Types of Microorganism and their differentiating characters | 5 |
| | | - Prokaryotes, Eukaryotes, three domain and five domain system | |
| | | of classification – | |
| | • | Bacteria (Eubacteria and Archaebacteria) | |
| | • | Protozoa | |
| | • | Fungi | |
| | • | Algae | |
| | • | Viruses, Viroids, Prions and Actinomycetes. | |

| II | Basic Techniques in Microbiology | 15 |
|----|--|----|
| | 1. Microscopy- | 5 |
| | 1.1 - Bright field microscopy [BFM]: | |
| | Electromagnetic spectrum of light | |
| | • Structure, working and ray diagram of a compound light microscope; | |
| | concepts of magnification, numerical aperture and resolving power. | |
| | • Types, ray diagram and functions of – Condensers (Abbe and cardioid) | |
| | eyepieces and objectives | |
| | • Concept of aberrations in lenses - spherical, chromatic, comma and | |
| | astigmatism | |
| | • Units of measurements of microorganisms | |
| | 2. Staining Techniques: | 5 |
| | • Definition of Stain; Types of stains (Basic and Acidic), | |
| | • Properties and role of Fixatives, Mordants, Decolourisers and | |
| | Accentuators. | |
| | Monochrome staining and Negative (Relief) staining | |
| | • Differential staining - Gram staining | |
| | • Special staining- Capsule, Spore | |
| | 3. Sterilization and Disinfection- | 5 |
| | • Sterilization: Physical Agents - Heat, Radiation, Filtration. | |
| | • Disinfection: Chemical agents and their mode of action – | |
| | Aldehydes, Halogens, Quaternary ammonium compounds, Phenol and | |
| | phenolic compounds, Heavy metals, Alcohol, Dyes, Detergents and | |
| | Ethylene oxide. | |
| | • Characteristics of an ideal disinfectant. | |

1. Atlas R. M. (1988, Digitized 2010). Experimental Microbiology: Fundamentals and Applications. United States: Macmillan.

2. Atlas R. M. (2005). Handbook of Media for Environmental Microbiology. United States: Taylor and Francis.

3. Atlas R. M. (2010). Handbook of Microbiological Media. Ukraine: Taylor and Francis.

4. Bergey's Manual of Systematic Bacteriology. (2005). Volume Two: The Proteobacteria, Part A: Introductory Essays. Garrity G. editor. Springer. ISBN 978-0-387-24143-2.

5. Bergey's Manual of Systematic Bacteriology. (2005). Volume Two: The Proteobacteria, Part B: The Gammaproteobacteria. Garrity G. Brenner D. J., Krieg N. R., and Staley J. R. (Eds.). Springer. ISBN 978-0-387-24144-9

6. Bergey's Manual of Systematic Bacteriology. (2005). Volume Two: The Proteobacteria, Part C: The Proteobacteria. Garrity G. Brenner D. J., Krieg N. R., and Staley J. R. (Eds.). Springer. ISBN 978-0-387-24145-6

7. Berry A. And Watson J. D. (2009). DNA: The Secret of Life. United States: Knopf Doubleday Publishing Group.

8. Bisen P.S. and Varma K. (2009). Handbook of Microbiology. CBS Publishers and Distributors, New Delhi. India

9. Biswas S. B. and Biswas A. (2006) Introduction to viruses. Vikas Publishing House Private Limited, New Delhi, India

10. Dubey H. C. (2004). A textbook of fungi, bacteria and Viruses. Vikas Publishing House Private Limited. New Delhi, India

11. Dubey R. C. and D. K. Maheshwari. (2012). A textbook of Microbiology. S Chand and Company. New Delhi, India

12. Gunasekaran P. (2007). Laboratory Manual in Microbiology. New Age International Private Limited, New Delhi, India.

13. Kapoor K. K., Tauro P. and Yadav K. S. (2016). An Introduction to Microbiology. New Age International (P) Limited, New Delhi, India.

14. Klein D. A., Harley J. P. And Prescott L. (2001). Microbiology. United Kingdom: McGraw Hill Higher Education.

15. Miller A. D. and Tanner J. (2013). Essentials of Chemical Biology: Structure and Dynamics of Biological Macromolecules. Germany: Wiley.

16. Parasher Y. K. (2006). Modern Microbiology. Campus Books International, New Delhi, India.

17. Pelczar M. J. Jr., Chan E.C.S. and Krieg N. R. (2010). Microbiology: An applicationbased Approach. McGraw-Hill Education (India) Private Limited, New Delhi, India.

 Powar C. B. and Daginawala H. I. (2005). General microbiology Volume I. Himalaya Publishing House Private Limited, Pune, India.

19. Powar C. B. and Daginawala H. I. (2005). General microbiology Volume II. Himalaya Publishing House, Private Limited, Pune, India.

20. Purohit S. S. (2006). Microbiology: Fundamentals and Applications. AGROBIOS, Jodhpur, Rajasthan, India.

21. Roush R., Wise J. and Fowler, S. (2018). Concepts of Biology. Hong Kong: Samurai Media Limited.

22. Singh R. P. (2012). Microbiology. Kalyani Publishers, New Delhi, India.

23. Singh R. P. and Singh B. D. (2009). Microbial Physiology and Microbial Genetics: (Microbiology). Kalyani Publishers, New Delhi, India.

24. Stanier R. Y. (2003). General Microbiology. United Kingdom: Palgrave Macmillan Limited.

25. Tortora G. J., Funke B. R. and Case C. L. (2016). Microbiology: an Introduction. Twelfth edition. Pearson, London.

26. Tsai C. S. (2007). Biomacromolecules: Introduction to Structure, Function and Informatics. Germany: Wiley.

 Yadav M. (2017). Microbiology. Discovery Publishing House, New Delhi, India. ISBN 13: 9788171417315.

Semester I Paper II MB-101-P - Laboratory Techniques in Microbiology I (2 Credits)

| Course Objectives | | |
|-------------------|--|--|
| 1 | To enrich students' knowledge and train them in core Microbiology. | |
| 2 | To provide students with hands-on experience in microbial staining techniques for microscopic examination. | |
| 3 | Students will get acquainted with skills of aseptic culture technique | |
| 4 | To introduce students to the techniques of microbial enumeration and quantification | |
| 5 | To make students proficient at laboratory skills and safety procedures. | |
| 6 | Students will develop critical thinking and problem-solving | |

| | Course Outcomes (COs) On completion of the course, the students will be able to: |
|-----|--|
| CO1 | Describe the good lab practices and biosafety measures to be adopted while working in a microbiology lab and identify different instruments commonly used for microbiological experiments. |
| CO2 | Give examples of different nutrient media popularly used in culturing microorganisms and compare different methods of sterilizing them. |
| CO3 | Demonstrate different cultivation methods for different microbial groups. |
| CO4 | Explain morphological characteristics of different microbial life forms by microscopic observation. |
| CO5 | Measure the dimensions of different biological cells. |
| CO6 | Design experiments to study the effect of sanitation methods on microflora |

1 Practical credit = 30 hours

1 Practical = 4 hours

| Expt. | Title | | |
|-------|--|---|--|
| No | | | |
| | i. Safety measures and Good Laboratory Practices in Microbiology | 3 | |
| | laboratory | | |
| 1 | ii. Introduction, operation, precautions and use of common | | |
| | Microbiology laboratory instruments: Incubator, Hot air oven, | | |
| | Autoclave, Colorimeter, Laminar air flow hood, Clinical centrifuge | | |
| | i. Construction (mechanical and optical), working and care of bright | 4 | |
| 2 | field microscope. | | |
| | ii. Permanent slide observation: Algae, Fungi and Protozoa | | |

| | iii. Wet mount preparation and its observation for: Bacteria, Algae, | |
|---|--|---|
| | Fungi and Protozoa | |
| | i. Introduction and use of common laboratory glassware: Test tubes, | 2 |
| | culture tubes, suspension tubes, screw capped tubes, Petri plates, | |
| 3 | pipettes (Mohr and serological) micropipettes, Pasteur pipettes, | |
| 5 | Erlenmeyer flask, volumetric flask, glass spreader, Durham's tube, | |
| | Cragie's tube and inoculating needles (wire loop, stab needles) | |
| | ii. Biological waste disposal (autoclaving) | |
| | Basic staining techniques: | 3 |
| 4 | i. Monochrome staining | |
| 4 | ii. Negative staining | |
| | iii. Gram staining of bacteria | |
| ~ | Observation of motility in bacteria using: Hanging drop and swarming | 2 |
| 5 | growth method | |
| | Checking of efficacy of chemical disinfectant: Determination of Phenol | 1 |
| 6 | Coefficient by Rideal–Walker method (demonstration) | |
| | Total | |

- Aneja K. R. (2007). Experiments in Microbiology, Plant Pathology And Biotechnology. New Age International, New Delhi, India
- Smith H. and Brown A. (2023). Benson's Microbiological Applications, Laboratory Manual,15th Edition. McGraw Hill.
- 3. Cappuccino J. G. and Welsh C. T. (2016). Microbiology: A Laboratory Manual. Pearson Education
- Deshmukh A. M. (2007). Handbook of Media Stains Reagents Microbiology. Oxford Book Company
- 5. Garratt D. C. (2012). The Quantitative Analysis of Drugs: 3rd Edition. United Kingdom:Springer US.

Semester I

Open Elective

OE-101-MB-Microorganisms and Human Life

Total: 2 Credits Workload:15hrs/credit (Total Workload:2 credits × 15 hrs =30 hrs in semester)

| Course Objectives This course is designed for students from non-microbiology background | | |
|---|---|--|
| 1 | Introduction to the invisible world of living organisms | |
| 2 | Understand distribution, diversity and differentiating characteristics of micoorganisms | |
| 3 | Awareness about use useful and harmful microorganisms | |

| | Course Outcomes After studying this course students will be able to |
|-----|---|
| CO1 | Understand concept of microorganism with respect to its size, shape, occurrence and types |
| | |
| CO2 | Differentiate between different types of microorganisms |
| CO3 | Understand principal and use of compound microscope and staining techniques to |
| | observe microorganisms |
| CO4 | Understand types of useful and harmful microorganisms |
| CO5 | Correlate the knowledge of microbiology in the day-to-day life for better living |

| Credit | Unit/ topic details | Number of Hours |
|--------|--|-----------------------|
| | 1. Introduction to Microbial World | 3 |
| | i. Discovery of microorganisms – Antony van Leuwenhoek | |
| | ii. Discovery of microscope – Leuwenhoek and Robert Hooke | |
| | iii. Diversity of microorganisms - Bacteria, Fungi, Algae, Protozoa, | |
| Ι | Viruses | |
| | iv. Types of bacteria (shapes and arrangements), morphological characteristics and habitat | |
| | 2. Microscopy | 1 |
| | • Principle and working of bright field microscope | |

| | 3. Staining of Microorganisms | | |
|----|--|---|--|
| | i. Purpose of staining | | |
| | ii. Monochrome staining | | |
| | iii. Negative staining | | |
| | 4. Cultivation of Microorganisms | 4 | |
| | i. Nutritional Classes of Microorganisms | | |
| | ii. Ingredients of Cultivation Media | | |
| | iii. Types of Media - Natural media (Coconut and Potato Slices) and | | |
| | Laboratory media (Nutrient Medium and Potato Dextrose Medium) | | |
| | 5. Sterilization and Disinfection | 5 | |
| | i. Sterilization – Definition and Agents (Physical and Chemical) | | |
| | ii. Pasteurization – Definition and Methods (Principle and | | |
| | Applications) | | |
| | iii. Disinfection - Definition and Chemical Agents | | |
| | 6. Beneficial and Harmful Effects of Microorganisms: | | |
| | a. Medical Microbiology (Vaccines - BCG & MMR and Antibiotics - | 3 | |
| | Pencillin & Streptomycin) | | |
| | b. Environmental Microbiology (Sewage Treatment and Bioremediation) | 3 | |
| II | c. Food and Dairy Microbiology (Food spoilage, Food-borne Diseases, | 3 | |
| | Probiotics and Fermented foods) | | |
| | d. Agriculture Microbiology (Plant Diseases and Bioinoculants) | 3 | |
| | e. Industrial Microbiology (Overview on Production of Antibiotics and | 3 | |
| | Enzymes) | | |

- Ananthnarayanan, R and Jeyaram Panicker, C. K. (2010). Textbooks of Microbiology. Orient Longman. 17th edition.
- Madigan, M.T., Martinko, J.M., Dunlap, P.V. and Clark, D.P. (2009).Brock Biology of Microorganisms, 12th edition, Pearson International edition, Pearson Benjamin Cummings.
- 3. Pelczar M. J. Jr., Chan E.C.S. and Krieg N. R. (2010). Microbiology: An Application based Approach. McGraw-Hill Education (India) Private Limited, New Delhi, India.
- 4. Willey, J. M., Sherwood L.M and Woolverton C.J., Prescott, (2013). Harley and Klein's. Microbiology. McGraw Hill Higher education. 9th Edition.

Semester I

Skill Enhancement Course [Practical] SEC-101-MB -Basic Skills in Microbiology Laboratory I

Total: 2 Credits Workload: 30hrs/credit

(Total Workload:2 credits \times 30 hrs =60 hrs in a semester) **1 Practical = 4 hours**

| This | Course Objectives This Course is designed to | | |
|------|--|--|--|
| 1 | Develop and enhance basic skills needed in microbiology laboratory for the beginners | | |
| 2 | Get hands on and job specific training of various instruments, glassware, chemicals and reagents used in the microbiology laboratory | | |
| 3 | Become more capable, competent, and confident in their performance | | |

| After | Course Objectives After studying this topic students will be able to. | | |
|-------|---|--|--|
| CO1 | Understand principle, working and calibration of various instruments used in microbiology laboratory and their SOPs | | |
| CO2 | Perform aseptic transfer of cultures | | |
| CO3 | Differentiate between different forms of microorganisms depending upon their microscopic and cultural characteristics | | |
| CO4 | Able to cultivate microorganisms on various natural and laboratory media | | |
| CO5 | Apply their knowledge and skills in clinical, pathological, food testing, environmental testing, etc. | | |

| Practical No. | Experiment Title | No. of Practicals |
|------------------|--|----------------------|
| 1 | Cleaning of laboratory glassware | 1 |
| 2 | Aseptic transfer techniques: Broth, Slant, Butt, Plate | 1 |
| 3 | Checking efficiency of autoclave (Chemical indicator) | 1 |
| 4 | Preparation of percent solutions (Volumetric flask) | 1 |

| 5 | Enumeration of yeasts using improved Neubauer chamber | 1 |
|----|---|----|
| 6 | Calibration of pH meter & checking of pH of natural samples - Fruit juices, milk, nutrient broth, distilled water, rain water, etc. | 1 |
| 7 | Preparation of laboratory media – Milk agar and potato dextrose agar (PDA) | 1 |
| 8 | Growth of microorganisms on natural surfaces – coconut, potato, bread | 1 |
| 9 | Cultivation and observation of protozoa – Hay infusion broth | 1 |
| 10 | Cultivation and observation of microorganisms form fermented foods (Idli batter, curd) through 100x objective – Monochrome staining | 2 |
| 11 | Check motility using Craige's tube method | 1 |
| 12 | Lipid granule staining | 1 |
| 13 | Demonstration of antibiotic sensitivity by disc diffusion method | 1 |
| 14 | Learning basic techniques in microbiology: Wrapping of glassware, making cotton plugs and plugging different glassware, cleaning and washing of glassware | 1 |
| | TOTAL | 15 |

- 1. Baunthiyal M., Saxena J. and Ravi I. (2015). Laboratory Manual of Microbiology, Biochemistry and Molecular Biology. Scientific Publishers, Jodhpur, Rajasthan, India.
- 2. Bisen P. S. (2014). Laboratory Protocols in Applied Life Sciences. United Kingdom: CRC Press.
- 3. Cappuccino J. and Welsh C. (2019). Microbiology: A Laboratory Manual, Loose Leaf Edition. United Kingdom: Pearson Education.
- 4. Dubey R. C. and Maheshwari D. K. (2012). Practical Microbiology. S. Chand and Company Limited, New Delhi, India
- 5. Gunasekaran P. (2007). Laboratory Manual in Microbiology. New Age International Private Limited, New Delhi, India.

Semester -II

Paper I

MB-151-T – Basics of Microbiology

Total: 2 Credits Workload:15 hrs/credit (Total Workload:2 credits × 15 hrs =30 hrs in semester)

| | Course Outcome After studying this course students will be able to |
|----|--|
| 1. | Understand the structure and functions of bacterial cells. |
| 2. | Acquire basic knowledge of biochemistry. |
| 3. | Develop an understanding of structure-function relationships in bio-molecules. |
| 4. | Learn the basics of the classification of bacteria and viruses |
| 5 | Understand, categorize, and differentiate nutritional classes, of microorganisms and |
| | apply them for nutrition cultivation, isolation, and preservation of different |
| | microorganisms |
| 6 | Relate, and apply knowledge of the life cycle stages of bacteria and growth behavior |
| | under different environmental conditions with various fields like fermentation |
| | technology, genetics, metabolism, pathogenesis, etc. |

| Credits | Unit/ Topic | No. of Hours (30) |
|---------|---|-------------------------|
| | 1. Bacterial Cytology | 10 |
| | Morphology -[Cell size, shape and arrangements of Bacteria] | |

| comp | onents in the bacterial cell: | |
|--|---|---|
| a. | Cell wall (Gram positive, Gram negative) | |
| b. | Concept of Mycoplasma, Spheroplast, protoplast, L-form | |
| c. | Cell membrane | |
| d. | Endospore (spore formation and stages of sporulation) | |
| e. | Capsule | |
| f. | Flagella | |
| g. | Fimbriae and Pili | |
| h. | Ribosomes | |
| i. | Chromosomal & extra-chromosomal material | |
| j. | Cell inclusion bodies. | |
| | Atom, Biomolecules, types of bonds (covalent, co-ordinate bond, non-covalent) and linkages (ester, phospho-diester, | |
| b. | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules | |
| | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates | |
| | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Proteins | |
| | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Proteins iii) Lipids | |
| b. | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Proteins iii) Lipids iv) Nucleic acids | |
| b. Classif | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Proteins iii) Lipids iv) Nucleic acids fication of Bacteria: | 1 |
| b. Classif | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Proteins iii) Lipids iv) Nucleic acids | 1 |
| b. Classif In Chara | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Proteins iii) Lipids iv) Nucleic acids fication of Bacteria: troduction to Bergey's Manual of Determinative Bacteriology | |
| b. Classif Int Chara 1. Cult | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Carbohydrates iii) Proteins iii) Lipids iv) Nucleic acids fication of Bacteria: troduction to Bergey's Manual of Determinative Bacteriology cteristics & Classification of Viruses: ICTV nomenclature | 1 |
| b. Classif Int Chara I. Cult a. 1 | bond, non-covalent) and linkages (ester, phospho-diester, peptide and glycosidic) Biomolecules: Classification, Structure, organization and functions of the following bimolecules i) Carbohydrates ii) Carbohydrates iii) Proteins iii) Lipids iv) Nucleic acids fication of Bacteria: troduction to Bergey's Manual of Determinative Bacteriology cteristics & Classification of Viruses: ICTV nomenclature tivation of Microorganisms: | 1 |

| c. Methods for cultivating photosynthetic, extremophilic, | |
|---|---|
| and chemo-lithotrophic bacteria, anaerobic bacteria, | |
| algae, fungi, actinomycetes, and viruses. | |
| d. Concept of Enrichment, Pure Culture, Isolation of culture by | |
| streak plate, pour plate, spread plate. | |
| e. Maintenance of bacterial and fungal cultures using different | |
| techniques. | |
| f. Culture collection centers and their role Requirements and guidelines of the National Biodiversity Authority for culture collection centers. | |
| 2. Bacterial growth: | 8 |
| a. Kinetics of bacterial growth (Exponential growth model) | 0 |
| b. Growth curve and Generation time | |
| c. Diauxic growth | |
| d. Measurement of bacterial growth- Methods of enumeration: | |
| i. Microscopic methods (Direct microscopic count, | |
| counting cells using improved Neubauer, Petroff- | |
| Hausser's chamber) | |
| ii. Plate counts (Total viable count) | |
| iii. Turbidometric methods (including Nephelometry) | |
| iv. Estimation of biomass (Dry mass, Packed cell volume) | |
| v. Chemical methods (Cell carbon and nitrogen estimation) | |
| e. Factors affecting bacterial growth [pH, Temperature, Solute | |
| e. Concentration (Salt and Sugar)] and Heavy metals(oligodynamic action) | |

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Semester -II

MB-151-P: Laboratory Techniques in Microbiology II

Total: 2 Credits Work-load:-30hrs/credit

(Total Workload:-2 credits x 30hrs =60 hrs in semester) **1 Practical = 4 hours**

| | Course Outcome After Completion of this course students will be able to | | |
|------|--|--|--|
| CO1. | Learn techniques of the cultivation of various types of organisms including skin microflora | | |
| CO2. | Understand the methods of bacterial enumeration from food, water or soil samples | | |
| CO3. | Acquainted and correlate the effects of various parameters including pH, salt concentration, temperature and heavy metal on bacterial growth | | |

| Expt | Topics | No. of |
|------|---|------------|
| No. | | Practicals |
| 1 | i. Preparation of simple laboratory nutrient media (Nutrient | 1 |
| | agar/broth, MacConkey's agar). | |
| | ii. Checking sterilization efficiency of autoclave using a | 1 |
| | biological indicator (B. stearothermophilus) | |
| | iii. Preparation of Winogradsky's column and observation of | 1 |
| | different types of microorganisms using bright field | |
| | microscope. | |
| 2 | Special staining techniques: | 2 |
| | i. Endospore staining | |
| | ii. Capsule staining | |
| 3 | Isolation of bacteria: | 1 |
| | Streak plate technique (Colony and cultural characteristics) | |
| 4 | Enumeration of bacteria from fermented food / soil / water by: | 2 |
| | i. Spread plate method | |
| | ii. Pour plate method | |

| 5 | Study of normal flora of skin: | 2 |
|---|---|----|
| | i. Cultivating and observing different morpho-forms of bacteria | |
| | from skin. | |
| | ii. Study of effect of washing on skin with soap and disinfectant on | |
| | it's microflora. | |
| 6 | To study the effect of different parameters on growth of <i>E. coli</i> : | 3 |
| | i. pH, temperature, sodium chloride concentration | |
| | ii. Study of oligodynamic action of heavy metal | |
| 7 | Preservation of cultures on: | 2 |
| | Slants, soil and on grain surfaces; revival of these cultures. | |
| | | |
| | TOTAL | 15 |
| | | |

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Semester II Open Elective OEP-151-MB -Dealing with Microorganisms

Total: 2 Credits Workload: 30hrs/credit

(Total Workload:2 credits \times 30 hrs =60 hrs in semester)1 Practical = 4.25 hours

| Course Objectives | | |
|-------------------|--|--|
| 1 | Students will get acquainted with skills of aseptic culture techniques | |
| 2 | To introduce students to the techniques of microbial production and quantification | |
| 3 | To make students proficient at laboratory skills. | |
| 4 | Students will develop critical knowledge about the domestic microbial processes | |

| Course Outcome After studying this course students will be able to | | |
|---|---|--|
| CO1 | Isolate and identify Nitrogen fixing bacteria from soil and leguminous plants and prepare biofertilizer from it | |
| CO2 | Perform primary screening of microorganisms of industrial importance | |
| CO3 | Develop skills for isolation and analysis of air & water bacteriologicaly | |
| CO4 | isolate and identify isolates from fermented foods | |
| CO5 | Analyse the results and interpret the observations | |

| Practical no. | Title | No. of Practicals |
|------------------|---|----------------------|
| 1 | Collection of various types of soils (activity-based) | 1 |
| 2 | Demonstration of Total Viable Count (TVC) of soil sample | 2 |
| 3 | Demonstration of antibiotic activity (Well-diffusion or disk-diffusion technique) | 2 |
| 4 | Microscopic observation of root nodule bacteria | 2 |
| 5 | Observation of air microflora | 2 |
| 6 | Demonstration of MPN for potability of water | 2 |
| 7 | Isolation of Lactic acid bacteria from fermented foods | 2 |
| 8 | Application of commercial bioinoculants (Pot study) | 2 |
| | Total | 15 |

- Aneja K. R. (2007). Experiments in Microbiology, Plant Pathology And Biotechnology. New Age International, New Delhi, India
- Cappuccino J. G. and Welsh C. T. (2016). Microbiology: A Laboratory Manual. Pearson Education
- Deshmukh A. M. (2007). Handbook of Media Stains Reagents Microbiology. Oxford Book Company
- Dubey R. C. and Maheswari D. K. (2000). Textbook of MicrobiologyS. Chand Publishing, New Delhi. ISBN: 9788121918039
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- Smith H. and Brown A. (2023). Benson's Microbiological Applications, Laboratory Manual,15th Edition. McGraw Hill.

Semester II

Skill Enhancement Course SEC-151-MB: Basic Skills in Microbiology Laboratory II

Total: 2 Credits Workload: 30hrs/credit

(Total Workload:2 credits \times 15hrs = 30 hrs in a semester)

| Course Objectives | |
|-------------------|--|
| 1 | To introduce students to the fundamental concepts and principles of microbiology and its |
| | practical applications. |
| 2 | To familiarize students with the basic laboratory techniques used in microbiology |
| | research and diagnostics. |
| 3 | To develop students' skills in aseptic technique and safe handling of microorganisms. |
| 4 | To enable students to understand and perform various methods of microbial cultivation |
| 5 | To teach students the principles and procedures involved in handling of instruments |

| | Course Outcomes After studying this course students will be able to |
|-----|--|
| CO1 | Handle compound microscope with objectives of different magnifications |
| CO2 | Isolate bacteria and yeasts from fermented foods |
| CO3 | Determine levels of contamination in various food products by counting methods |
| CO4 | Learn methods of isolation and identification of various plant pathogens |
| CO5 | Check antimicrobial activities of U.V light, various spices, condiments and herbs used in preparation of foods |
| CO6 | Record observations, analyse the data, interpret results and draw conclusions |

| Sr. No. of expt. | Experiment | No. of practicals |
|---------------------|---|----------------------|
| 1 | Microscopic observation of mouth flora by monochrome staining | 1 |
| 2 | Check antimicrobial activity of U.V. light & efficiency of HEPA filters | 1 |
| 3 | Staining of fungi (using lactophenol cotton blue) | 1 |

| 4 | Isolation of yeast from natural sample and staining | 2 |
|----|--|----|
| 5 | Determination of microbial load of milk – standard plate count | 1 |
| 6 | Isolation and observation of bacterial plant pathogens from soft rot of potato/ citrus fruit. | 1 |
| 7 | Observation of plant pathogens from. 1. Black rust of wheat 2. Leaf curl of tomato 3. Downy mildew of grape 4. Red rot of sugarcane 5. Whip smut of Sugarcane (any two of the above) | 2 |
| 8 | Demonstration of Koch's postulates | 1 |
| 9 | Determination of antibacterial activity of – clove, pepper, garlic, onion (well/ disc diffusion method) | 2 |
| 10 | Check pasteurization efficiency of milk by phosphatase test | 1 |
| 11 | Check sterility of pharmaceutical products – fluids & powders | 2 |
| | TOTAL | 15 |

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- Bisen P. S. (2014). Laboratory Protocols in Applied Life Sciences. United Kingdom: CRC Press.
- Dubey R. C. and Maheshwari D. K. (2012). Practical Microbiology. S. Chand and Company Limited, New Delhi, India
- Gunasekaran P. (2007). Laboratory Manual in Microbiology. New Age International Private Limited, New Delhi, India.
- Harley J. P. and Prescott L. (2020). Laboratory Exercises in Microbiology. Independently Published.
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- Sharma K. (2007). Manual of Microbiology (Second Edition). ANE Books, New Delhi, India.
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